

Intrinsically Disordered Protein, Alternative Splicing & Post-Translational Modification (IDP-AS-PTM): A Toolkit for Developmental Biology

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Textbook Protein Structure/Function

**Currently
Dominant
Protein
Structure/
Function
Paradigm**

Amino Acid Sequence

“Folding Problem”



3-D Structure

Native = Ordered = Structured



Protein Function

[“Lock & Key”; “Induced Fit”]

Definition: Intrinsically Disordered Proteins (IDPs) and IDP Regions

Whole proteins and regions of proteins are **intrinsically disordered** if:

- they lack stable 3D structure under physiological conditions, and if:
- they are flexible molecules that form dynamic ensembles with inter-converting configurations and without particular equilibrium values for their coordinates.

Intrinsically Disordered Proteins (IDPs)

- Karl Landsteiner (1939) & Linus Pauling (1940) suggested that **unfolded proteins** exist and that **they** fold into different **structures** as **they** bind separately to multiple, differently **shaped** partners.
- **IDPs** first characterized in the 1950s by **OR & ORD**.
- Thousands now characterized by **X-ray, NMR, etc.**, & especially by **computational biology & bioinformatics**.
- **IDP** discovery represents a **true paradigm shift**.

Reviewed in Dunker AK & Oldfield CJ *Adv Exp Med Biol* 870: 1-34 (2015)

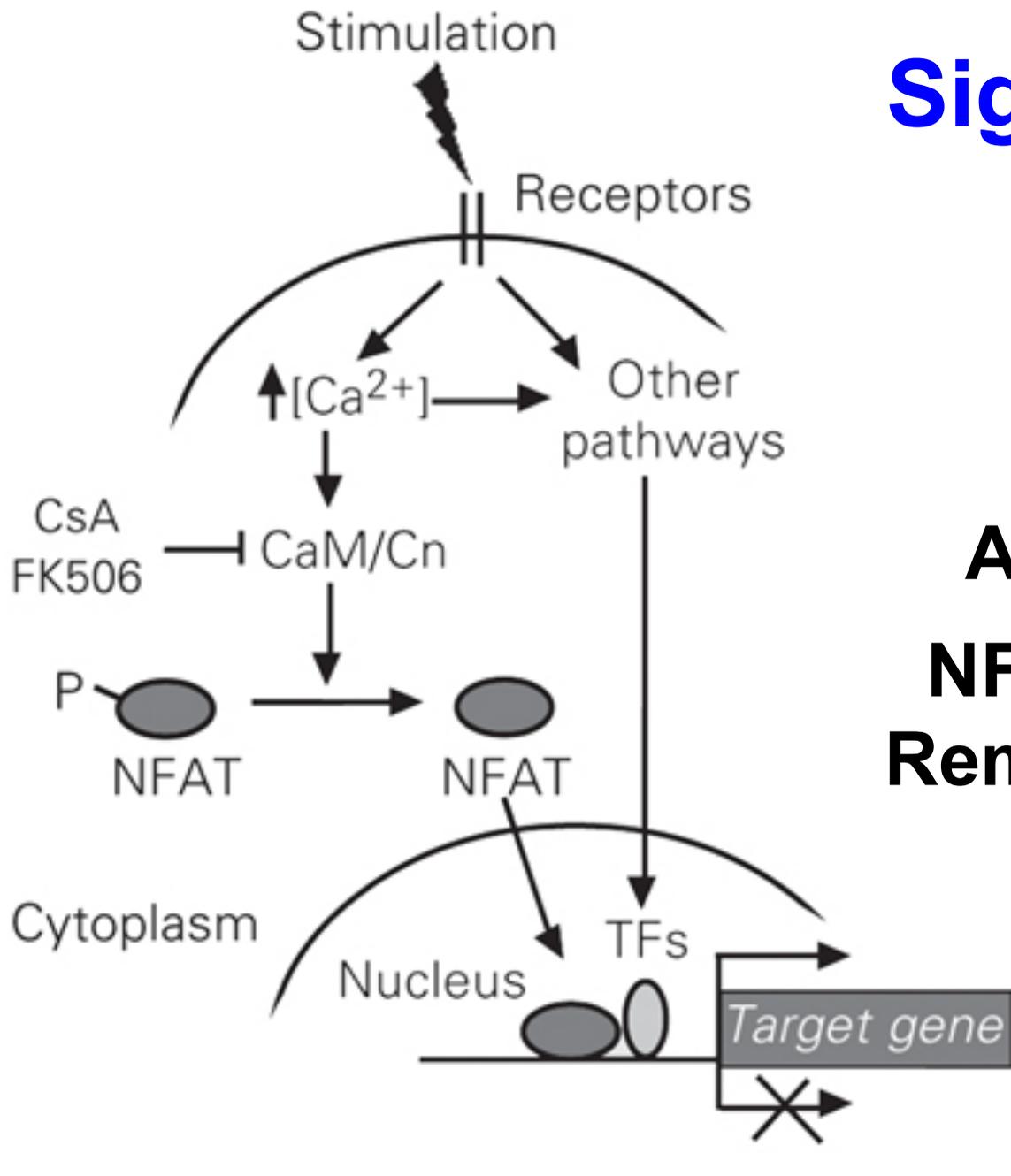
Trigger for Dunker's **IDP** Research



Seminar describing an important **IDP**
12 Noon to 1 PM, 15 November, 1995
Washington State University

Given By Chuck Kissinger
BS / MS Washington State University
PhD University of Washington
Johns Hopkins / MIT Post Doc
Aguoron Pharmaceuticals

Signaling Pathway



Calmodulin (CaM)
Calcineurin (Cn)
Nuclear Factor of Activated T- Cells (NFAT)

NFAT-poly-P in an IDP tail.
Remove Ps, activates NLS

→ NFAT → nucleus

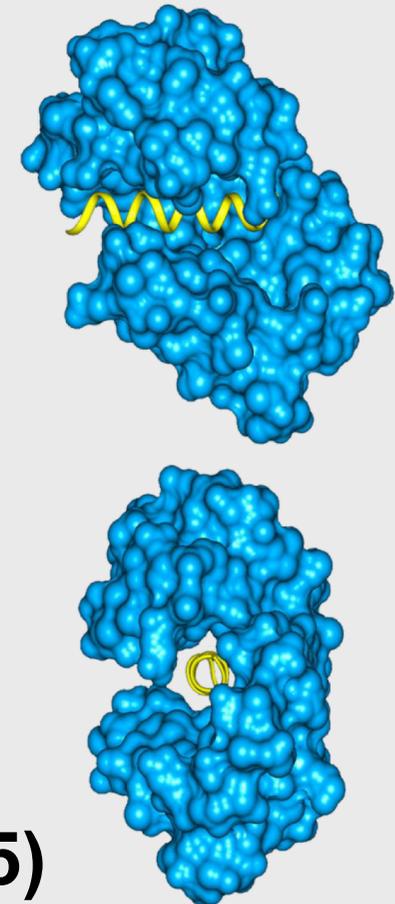
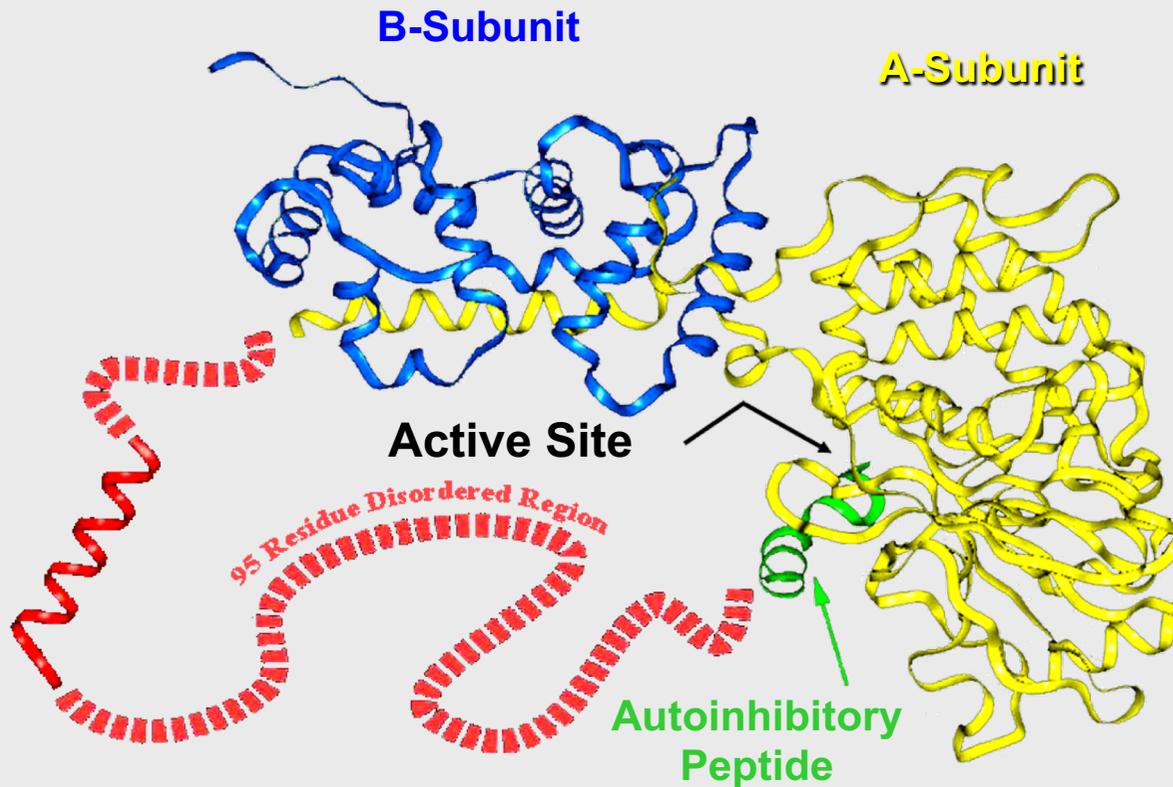
→ turns on genes

→ T-cells activated

→ reject transplant

Calcineurin and Calmodulin

Meador W et al., *Science*
257: 1251-1255 (1992)



Kissinger C et al., *Nature* 378:641-644 (1995)

Intrinsically Disordered Proteins (IDPs)

After Seminar Questions:

- Why don't **IDPs** and **IDP regions** fold into 3D **structure**?
- How common are **IDPs** and **IDP regions**?
- What are the functions of **IDPs** and **IDP regions**?

Why don't **IDPs** fold into **3D structure**?

- **Amino acid composition** determines whether a protein will **fold** or remain **unfolded**.
- For compositions that favor **structure**, the sequence patterns of hydrophobic / hydrophilic groups determine which **3D structure** is formed.

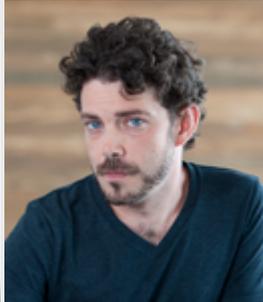
Shakhnovich, E.I. and Gutin, A.M. Engineering of stable and fast-folding sequences of model proteins. *Proc. Natl. Acad. Sci. USA* 90: 7195 – 7199 (1993).

Why don't **IDPs** fold into 3D **structure**?

Xie et al., *Genome Informatics* 9: 193-200 (1998)



Qian
Xie



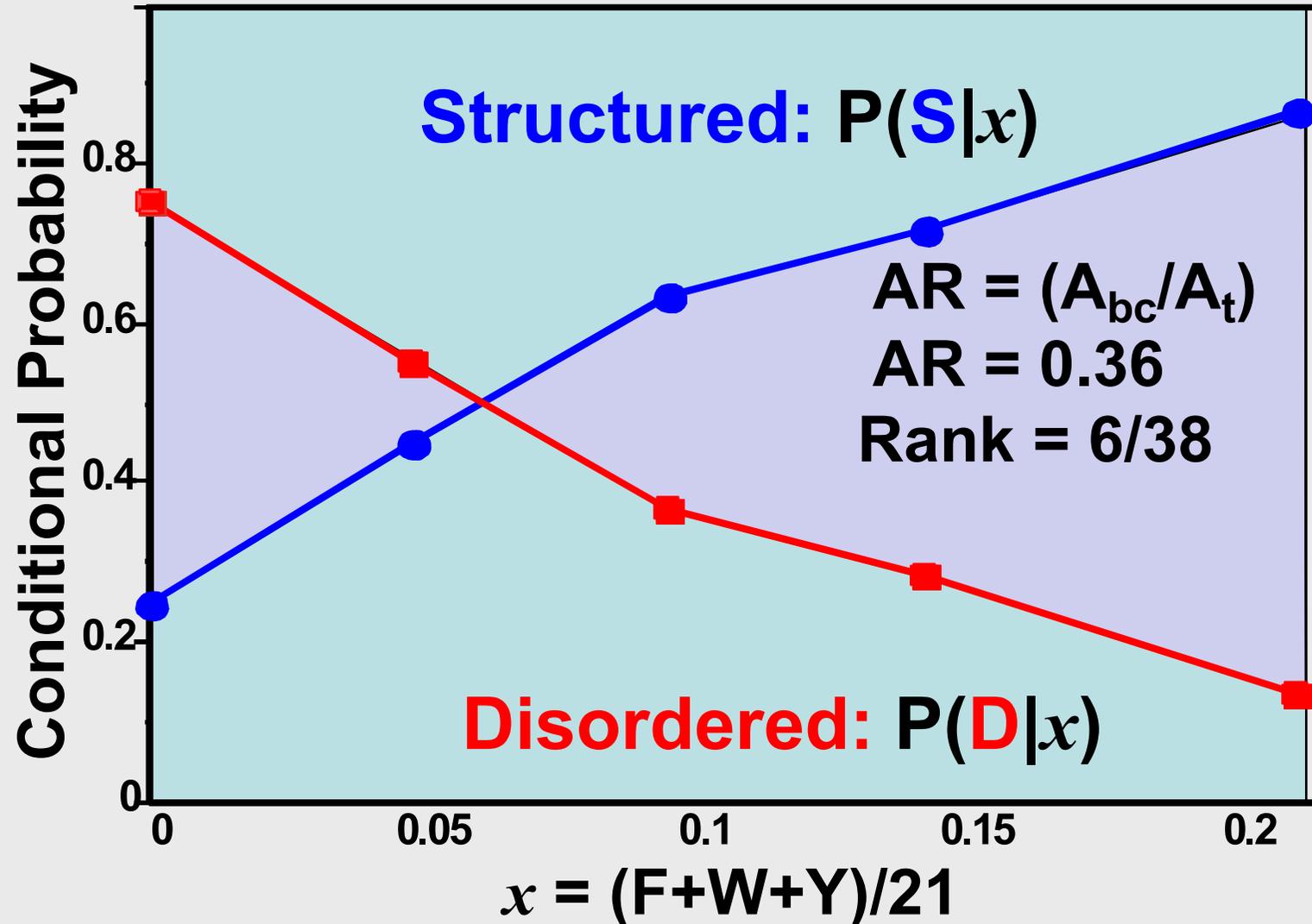
Ethan
Garner



Zoran
Obradovic



Pedro
Romero



Why don't **IDPs** fold into 3D **structure**?

Amino acid sequence favors nonfolding!

- **IDPs** have too few aromatics – aromatics are important for the stability of hydrophobic cores;
- **IDP** ratio of hydrophilic amino acids to hydrophobic amino acids is too high for folding;
- **IDPs** have too low of a sequence complexity
- **IDPs** have too large of a net charge – charge repulsion inhibits folding;
- **IDPs** have too many prolines – prolines cannot form backbone H-bond, so helices and sheets are destabilized by prolines.

Intrinsically Disordered Proteins (IDPs)

How common are **IDPs** and **IDP regions**?

Step 1: Develop predictor of **IDPs** and **IDP regions**.

Step 2: Apply to multiple **proteomes**.

Dunker et al., *Genome Informatics* 11: 161-171 (2000)
(repeated by many others, and by us)

Step1: Predictor **Intrinsic Disorder**

Disordered & Ordered Sequence Data

Attribute Selection or Extraction

Separate Training and Testing Sets

Predictor Training

Predictor Validation on Out-of-Sample Data

Prediction

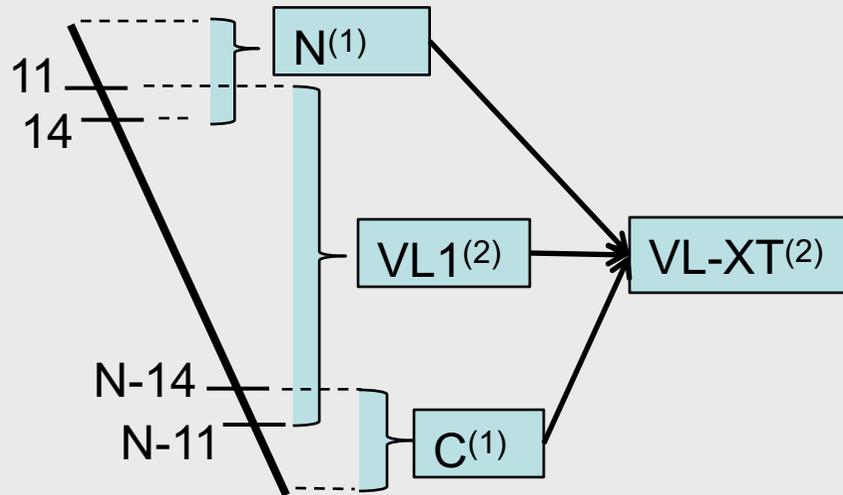
**Aromaticity,
Hydropathy,
Net Charge,
Complexity**

**Neural Networks,
SVMs, etc.**

**CASP Expt: 2002 – 2010
Bal. ACC ~ 0.75; AUC ~ 0.86**

Predictors of Natural Disordered Regions

PONDR[®]VL-XT and PONDR[®]VSL2

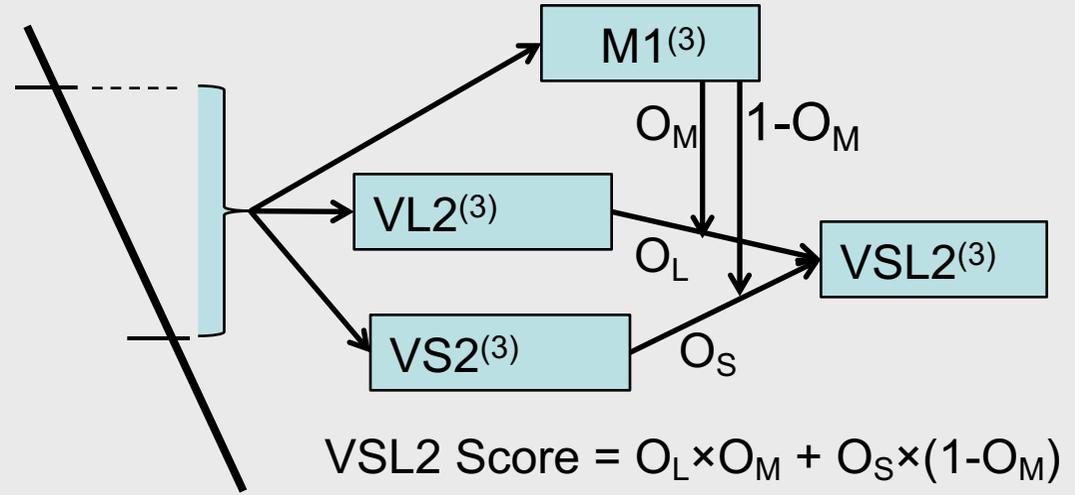


N, VL1, and C are neural networks

N-term: 8 inputs

VL1: 10 inputs

C-term: 8 inputs



$$\text{VSL2 Score} = O_L \times O_M + O_S \times (1 - O_M)$$

M1, VSL2-L, and VSL2-S are support vector machines

M1: 54 inputs

VL2: 20 inputs

VS2: 20 inputs

(1) Li X et al., *Genome Informat.* 9:201-213 (1999)

(2) Romero P et al., *Proteins* 42:38-48 (2001)

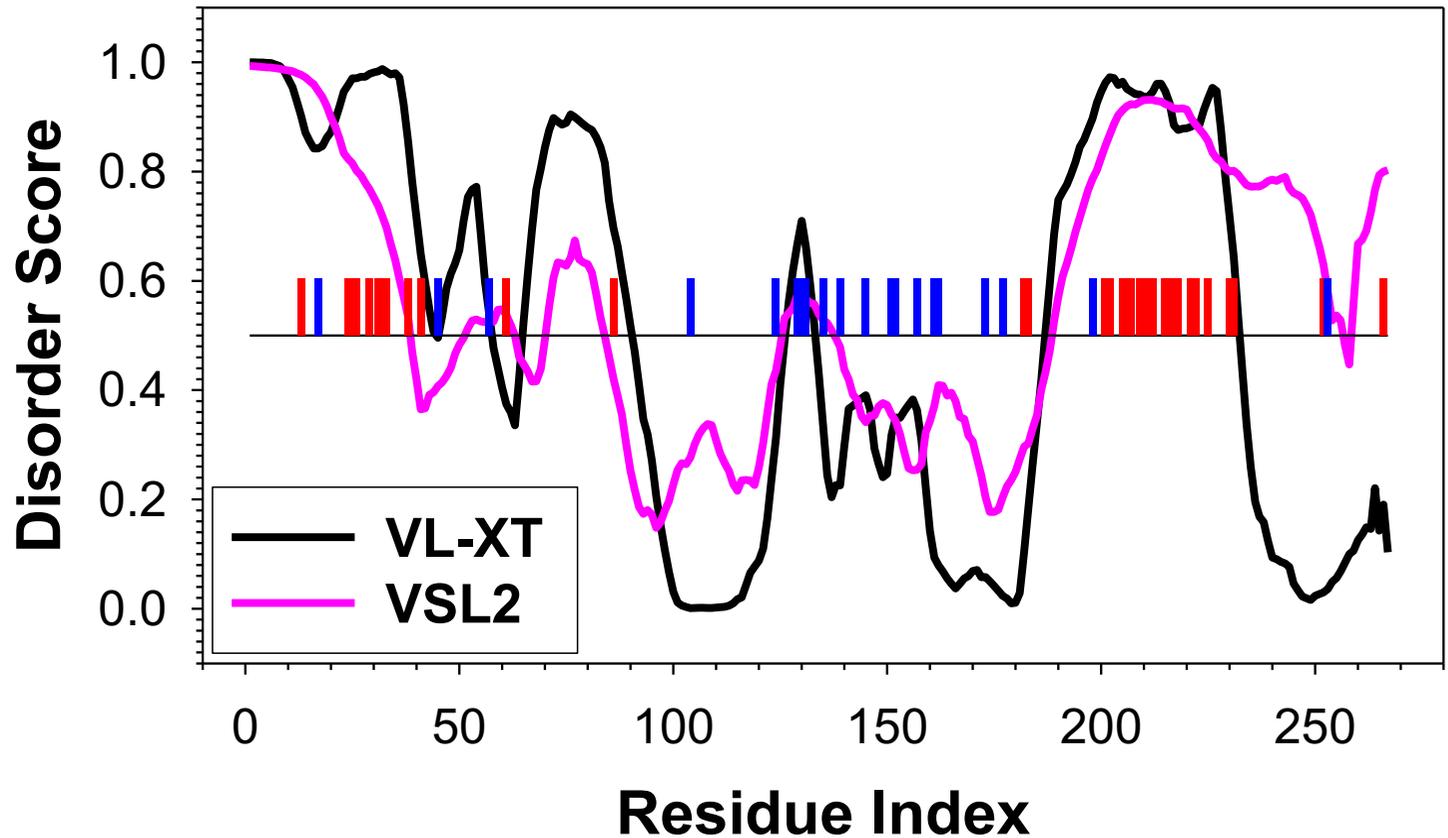
(3) Peng K et al., *BMC Bioinfo.* 7:208 (2006)

PONDR[®]VL-XT and PONDR[®]VSL2

(+) Disordered

XPA

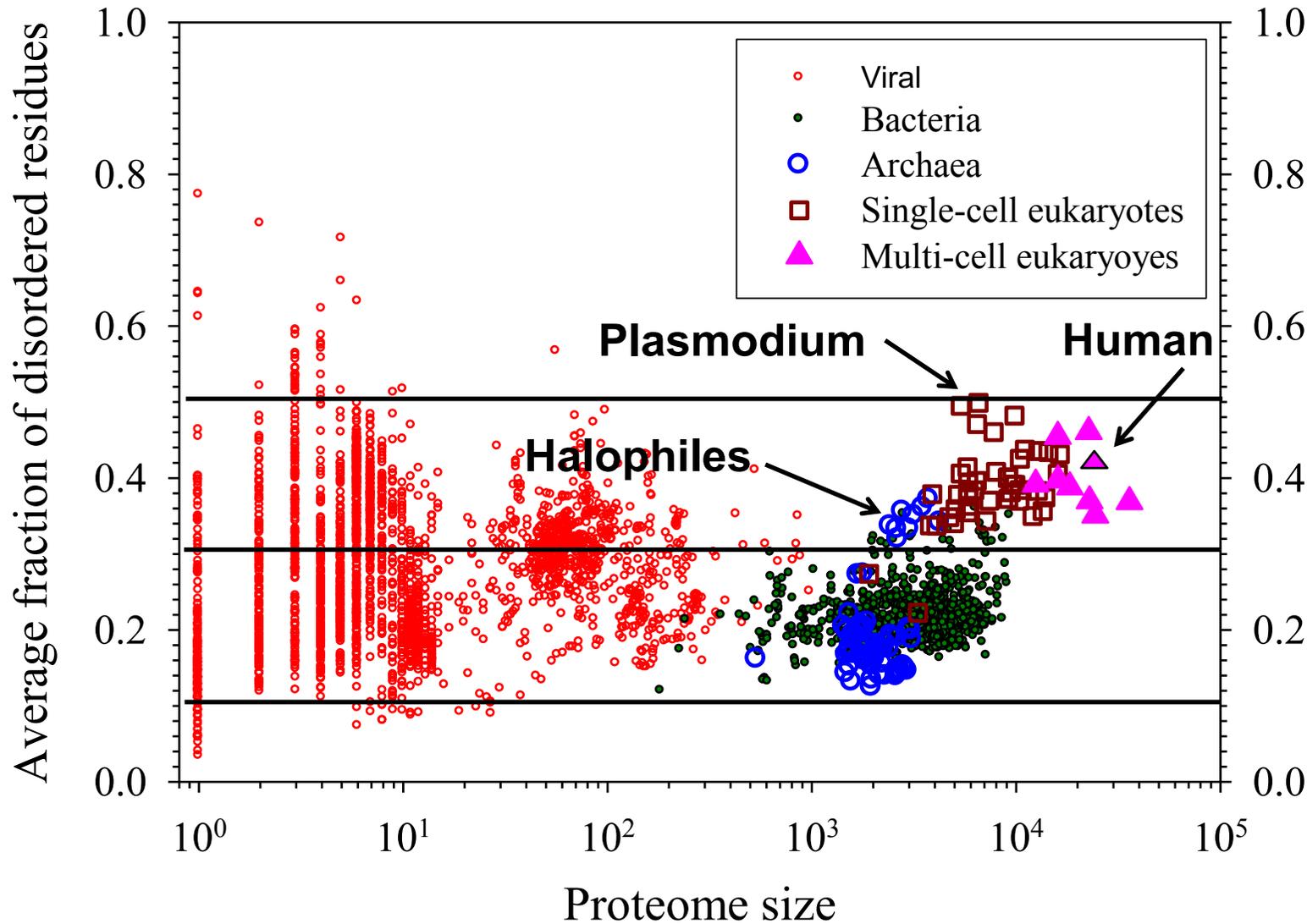
(-) Structured



Iakoucheva L et al., *Protein Sci* 3: 561-571 (2001)

Dunker AK et al., *FEBS J* 272: 5129-5148 (2005)

Step 2: How common are IDPs?



Bin Xue



Vladimir Uversky

Xue et al., *J Biomol Struct Dyn* 30: 137-149 (2012)

How common are **IDPs**?

More recent, improved approach

Combine **structure / disorder** prediction and **structure prediction** by *sequence similarity* to all currently known protein 3 D structures.

For the human proteome:

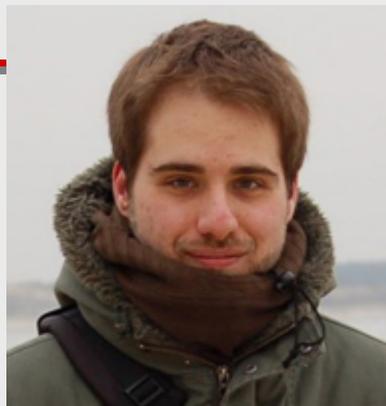
Fukuchi, S., *et al.*, Binary classification of protein molecules into **intrinsically disordered** and **ordered** segments. *BMC Struct Biol.* 11:29 (2011); For Human: 35% residues are in **IDPs** or **IDP regions**. (**Weakness** → used **Pfam** for **structured proteins**)

For 1,765 proteomes (8 different **order / disorder** predictors):

Oates, M.E. *et al.*, D²P²: database of disordered protein predictions. *Nucleic Acids Res.* 41(Database issue):D508-516 (2013). For Human: 35% - 50% residues in **IDPs** or **IDP regions**. (**Strength** → used **SUPERFAMILY** for **structured proteins**)

Human BIN1 from D²P²

Two transcripts from one gene;

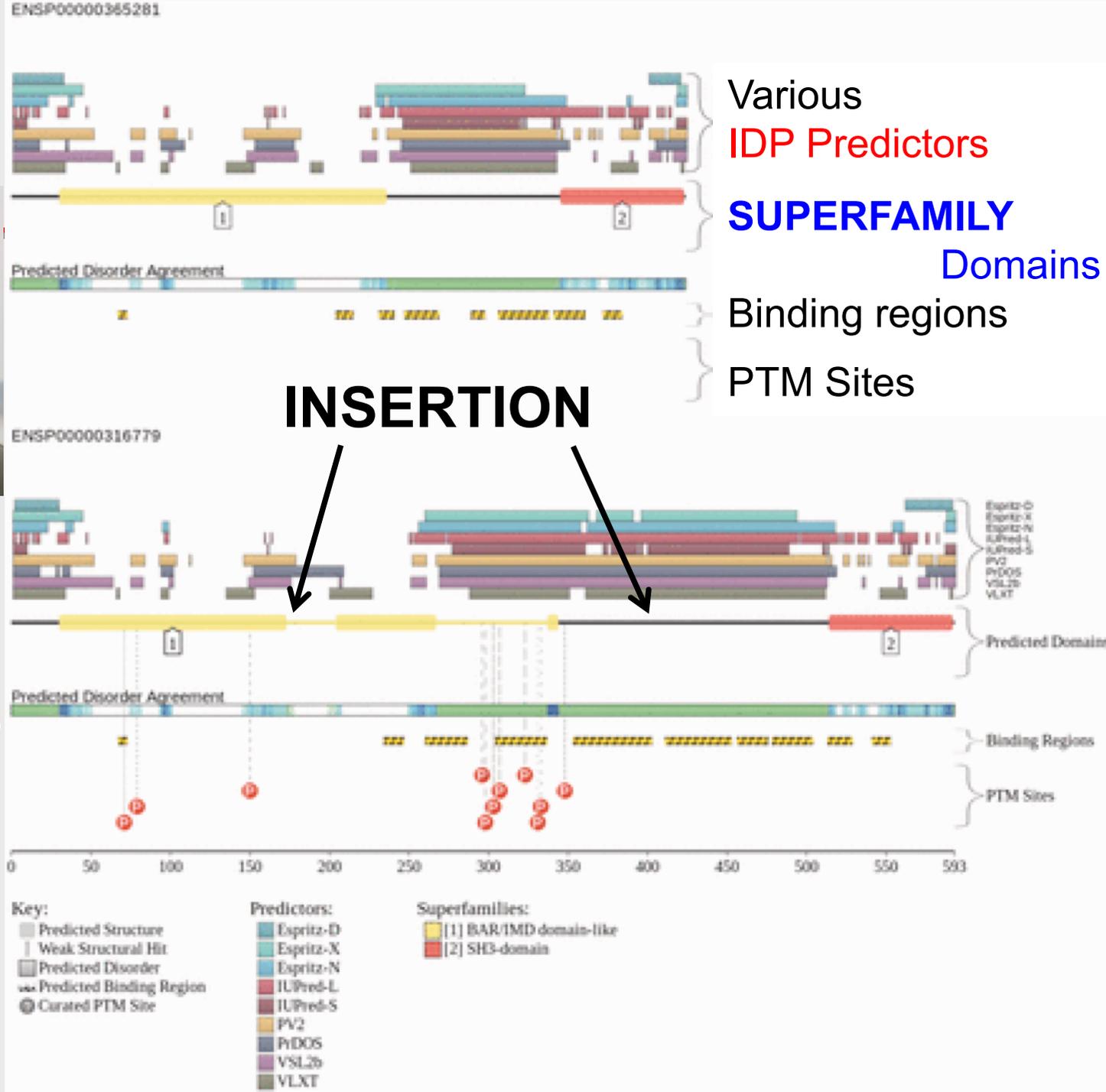


Matt Oates



Insertion from alternative splicing.

Julian Gough



Oates M *et al.*, *NAR*
41: D508-516 (2013)

IDP Functions: Lac Repressor

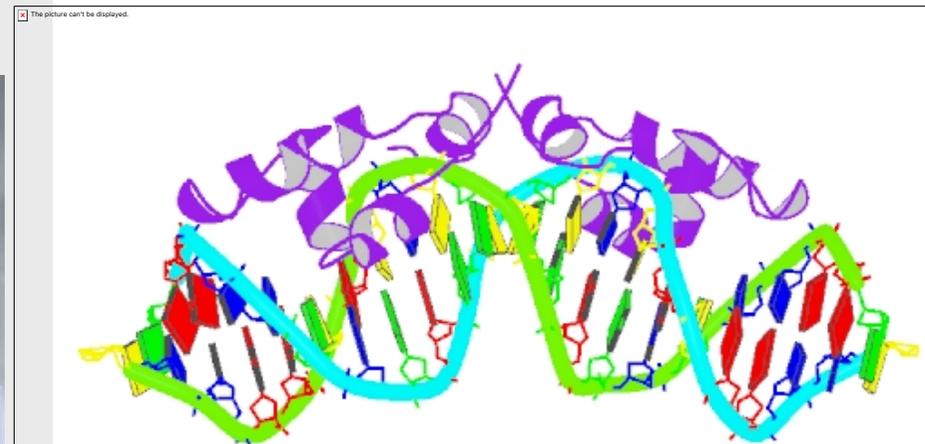
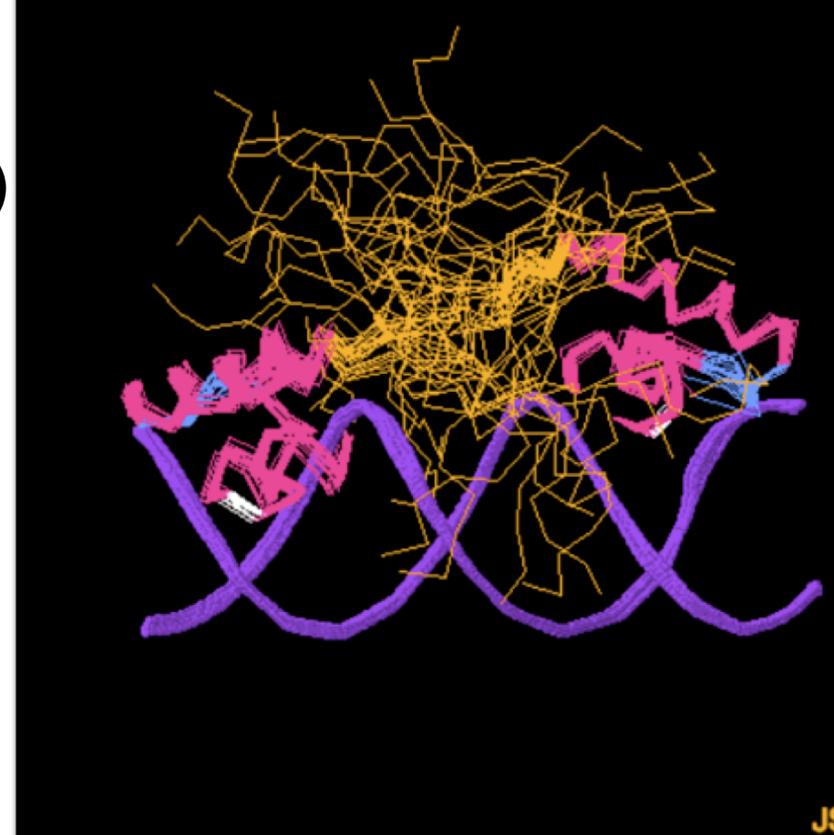
Kalodimos *et al.*, *Science* 305:386-389 (2004)

- Upon binding random DNA, a 12 residue linker remains **disordered** & binds DNA phosphates transiently, helping the **Lac Repressor** slide along the **DNA**.
- Upon encountering its binding sequence, the **IDP region** → **structure** and is involved in recognizing the **cognate DNA binding sequence**, in increasing the affinity, & in helping bend the **DNA**.

Images: **Proteopedia**, **Life in 3D**, the free, collaborative, 3D Encyclopedia:



– provided by: **Joel Sussman**



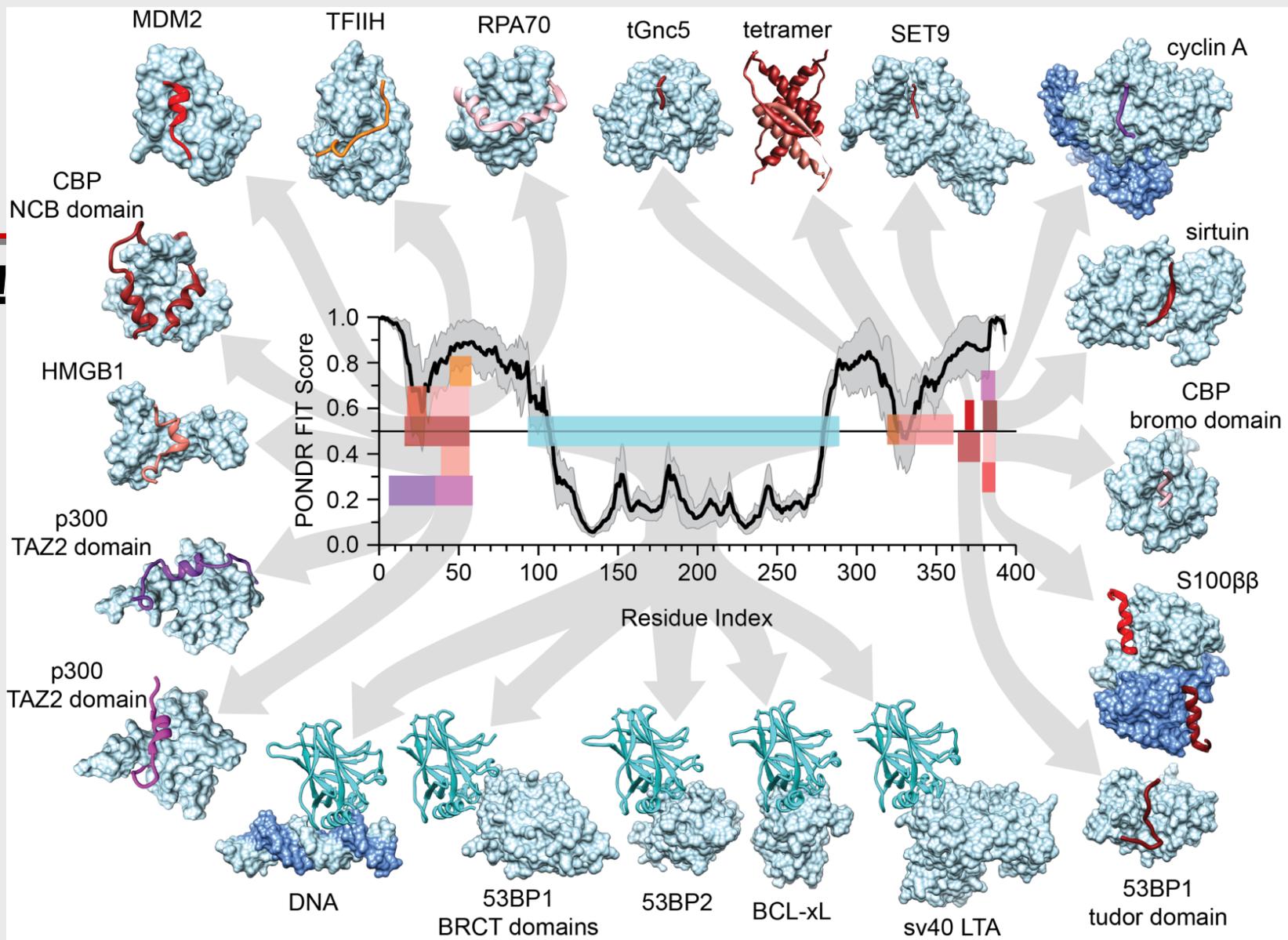
p53 binding

Note **IDP** tails!

Molecular Recognition Features (MoRFs)

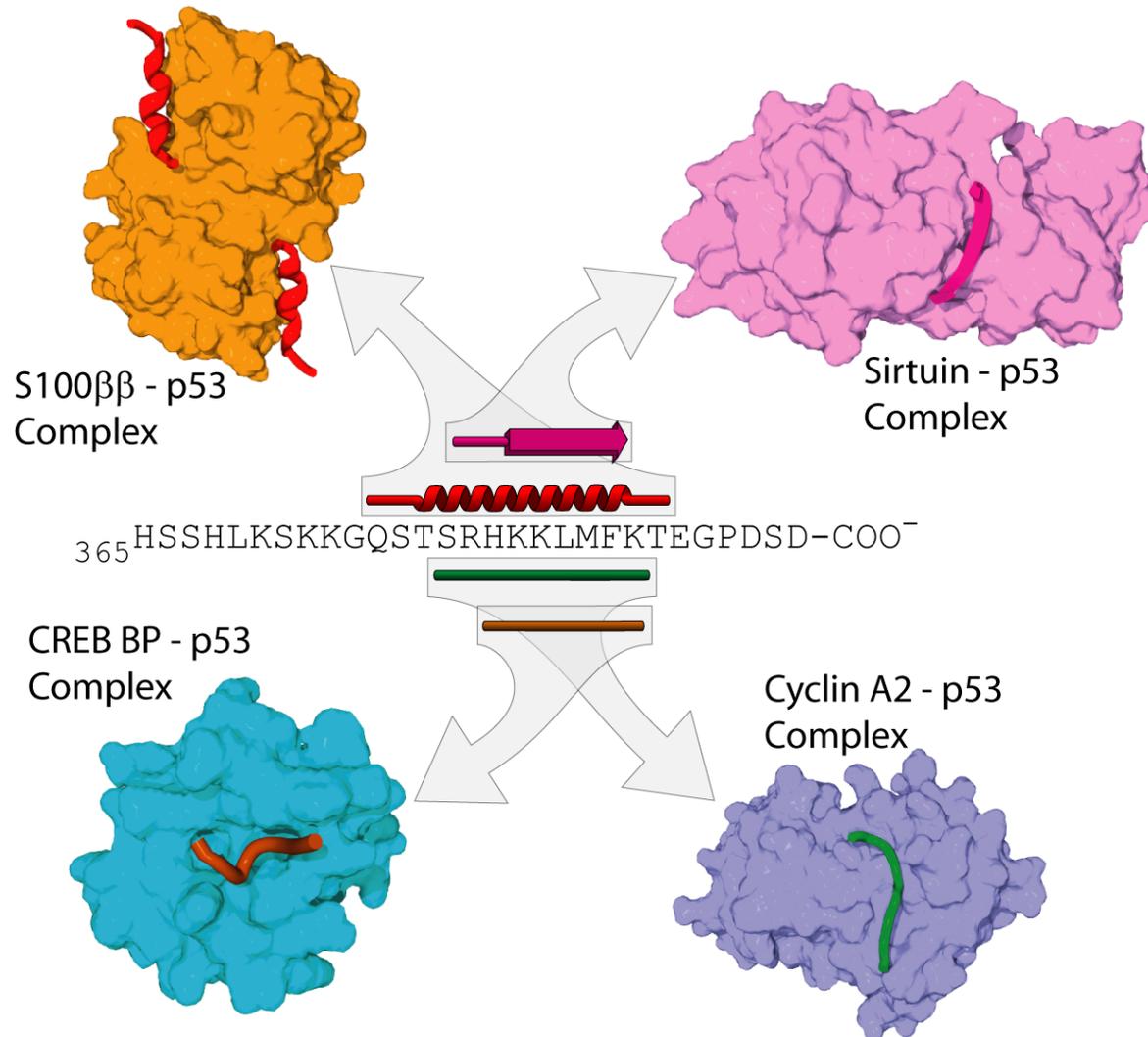


Chris Oldfield



Modified from: Oldfield & Dunker, *Ann Rev Biochem* 83: 553 – 584 (2014)

p53 C-terminal Domain: Secondary Structure and Overlap

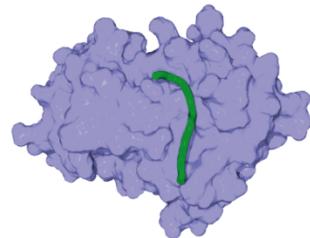
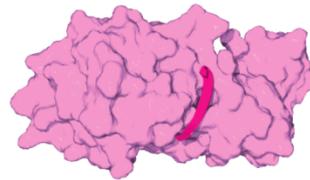
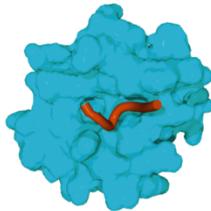
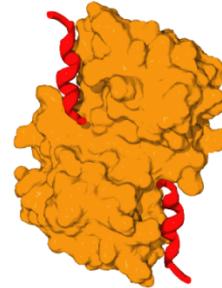
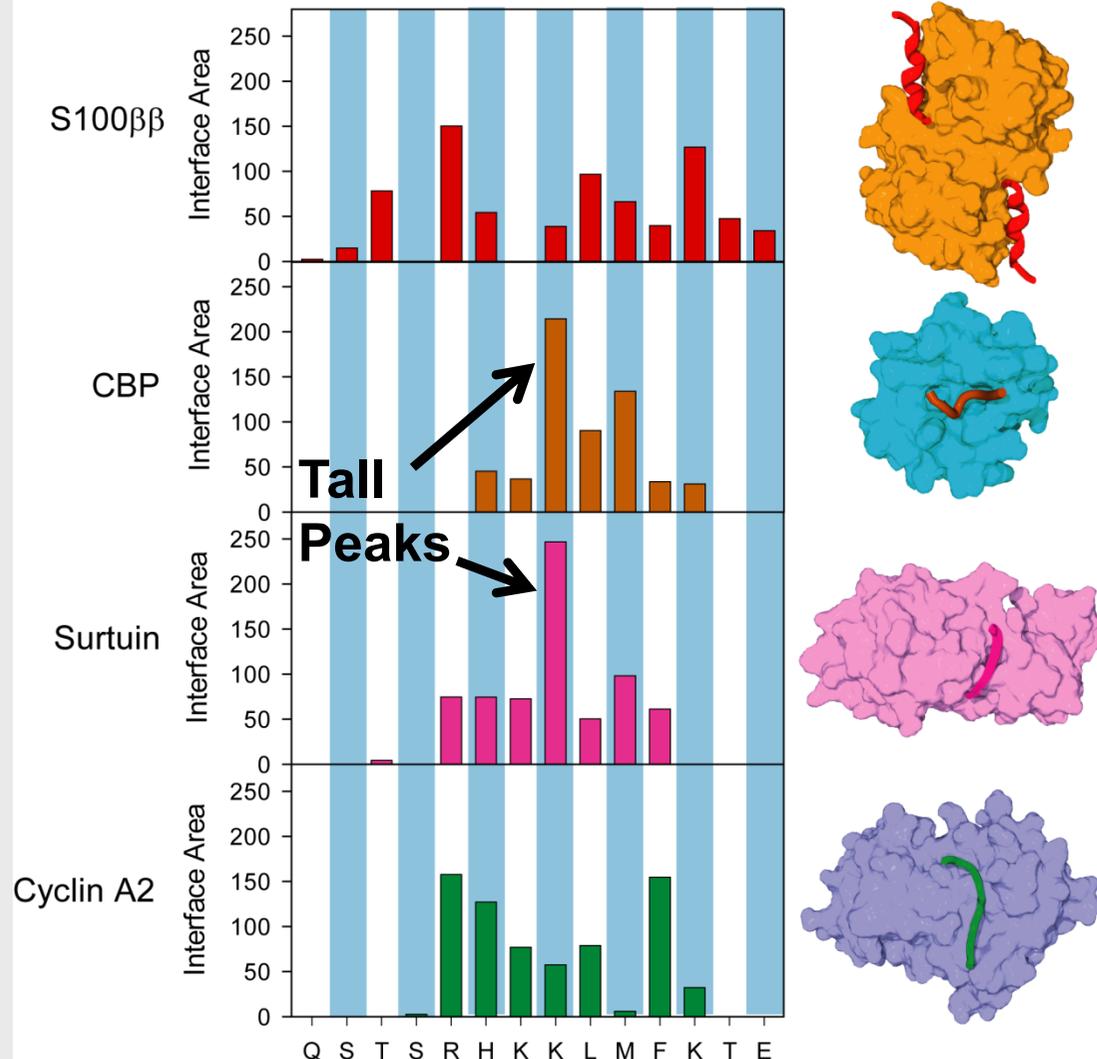


Oldfield CJ, et al.,
BMC Genomics
9 (Suppl 1) S1 (2008)

Confirms Landsteiner-Pauling 1939 - 1940 hypothesis of changes in structure due to folding upon binding to different partners!!

Pauling L, J Am Chem Soc
62: 2643-2657 (1940)

p53 C-terminal **IDP** region: Residue-specific Interface Area



Oldfield CJ, et al.,
BMC Genomics
9 (Suppl 1) S1 (2008)

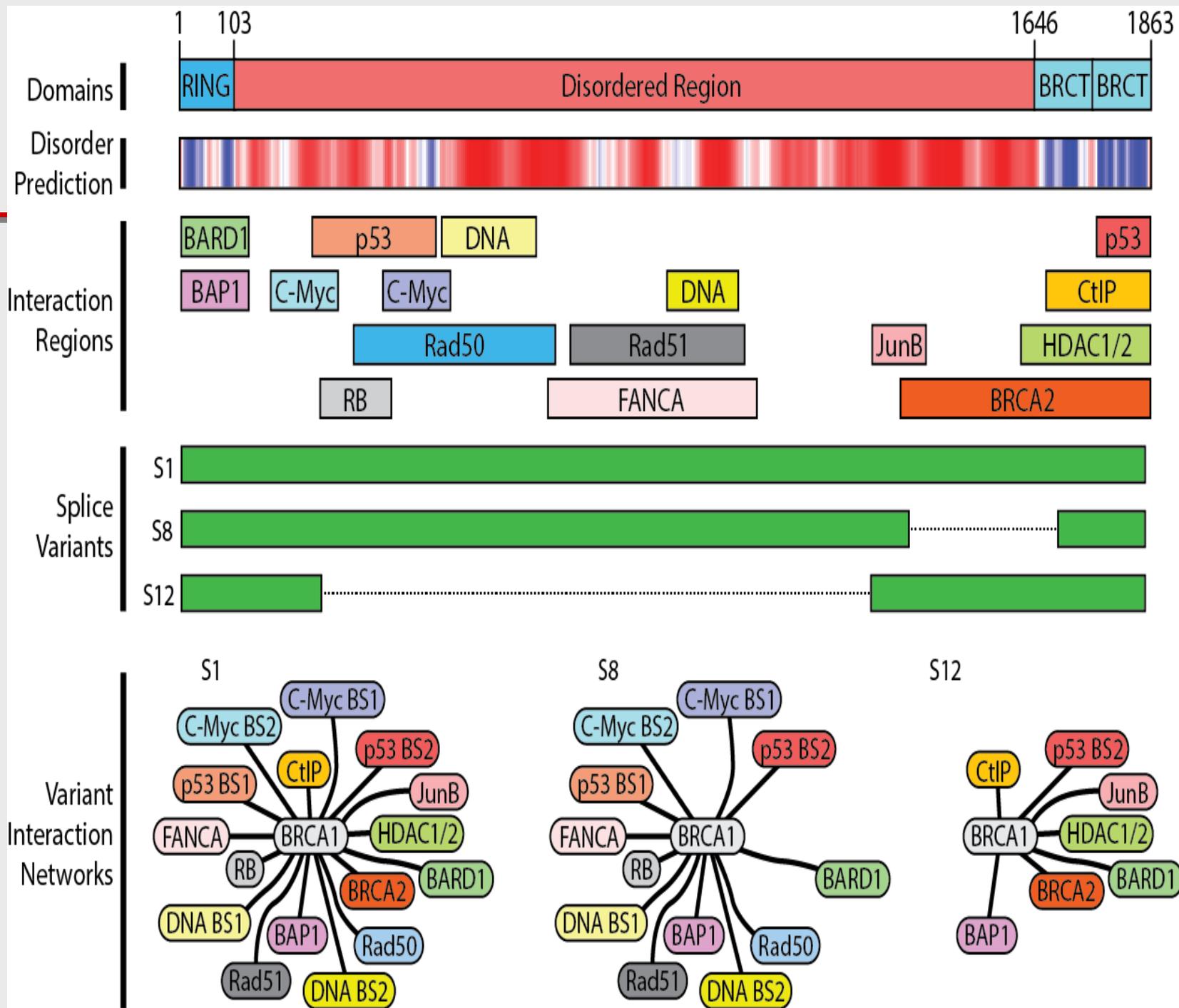
STSRHKKLMFKE

Tall peaks in CBP and Sirtuin; due to buried acetyl group.

PTMs often contribute to partner switching. Many examples observed.

BRCA1

1863
residues;
103 ordered
at N-term;
217 ordered
at C-term;
1543
disordered
in between.



The **IDP-AS-PTM** Toolkit Hypothesis

IDP, AS, & PTM all shown to enable signalling complexity:

- **IDPs change shape** & thereby **bind** to **multiple partners**.
- **PTMs** within **IDP regions** bring about **partner switching**.
- **AS** of mRNA coding for **IDP regions** **rewires** protein-protein & protein-DNA interaction networks – often **tissue-specific!**

Hypothesis: **IDP, AS, PTMs** are **colocalized** & thus **collaborate** to further increase signaling complexity.



Hongbao Xie

IDPs & Function Global Analysis



Zoran Obradovic

- Collect SwissProt function-specific sequences;
- Collect 1,000 matching, random-function sequences; Matching = same size, same # chains.
- Predict **disorder** for each function-specific & 1,000 random-function sets \rightarrow all RFS sets \sim Gaussian;
- Rank **structure**- and **disorder**-associated functions by Z-scores ($Z\text{-score} = [x - \langle x \rangle] / \sigma$); Set $\langle x \rangle = 0$.
 - values = more structure, + values = more disorder

IDPs & Function

Functional Key Word Categories	Number
High-prediction of disorder ($> +1$)	238
Intermediate (Z-score, -1 to $+1$)	170
Low-prediction of disorder (< -1)	302
TOTAL	710

Xie H et al. *J. Proteome Res.* 6: 1882- 1898;
6:1899-1916; & 6:1917-1932 (2007)

Top 10 **Biological Processes** Most Strongly Associated with Low-prediction of **Disorder** (e.g. with **Structure**)

KEYWORDS	Proteins (number)	Families (number)	Length (Ave)	Z – Score
<i>GMP Biosynthesis</i>	225	3	473	-17.6
<i>Amino-acid Biosynthesis</i>	7098	212	361	-17.1
<i>Transport</i>	19888	2199	378	-14.9
<i>Electron Transport</i>	4633	346	272	-13.7
<i>Lipid A Biosynthesis</i>	533	13	291	-13.2
<i>Aromatic Catabolism</i>	320	105	300	-12.4
<i>Glycolysis</i>	2255	50	390	-12.1
<i>Purine Biosynthesis</i>	1208	28	445	-11.9
<i>Pyrimidine Biosynthesis</i>	1310	27	383	-11.7
<i>Carbohydrate Metabolism</i>	1797	180	404	-11.7

Xie H, et al., *J. Proteome Res* 6: 1882-1932 (2007)

Top 10 **Biological Processes** Most Strongly Associated with High-Prediction of **Disorder**

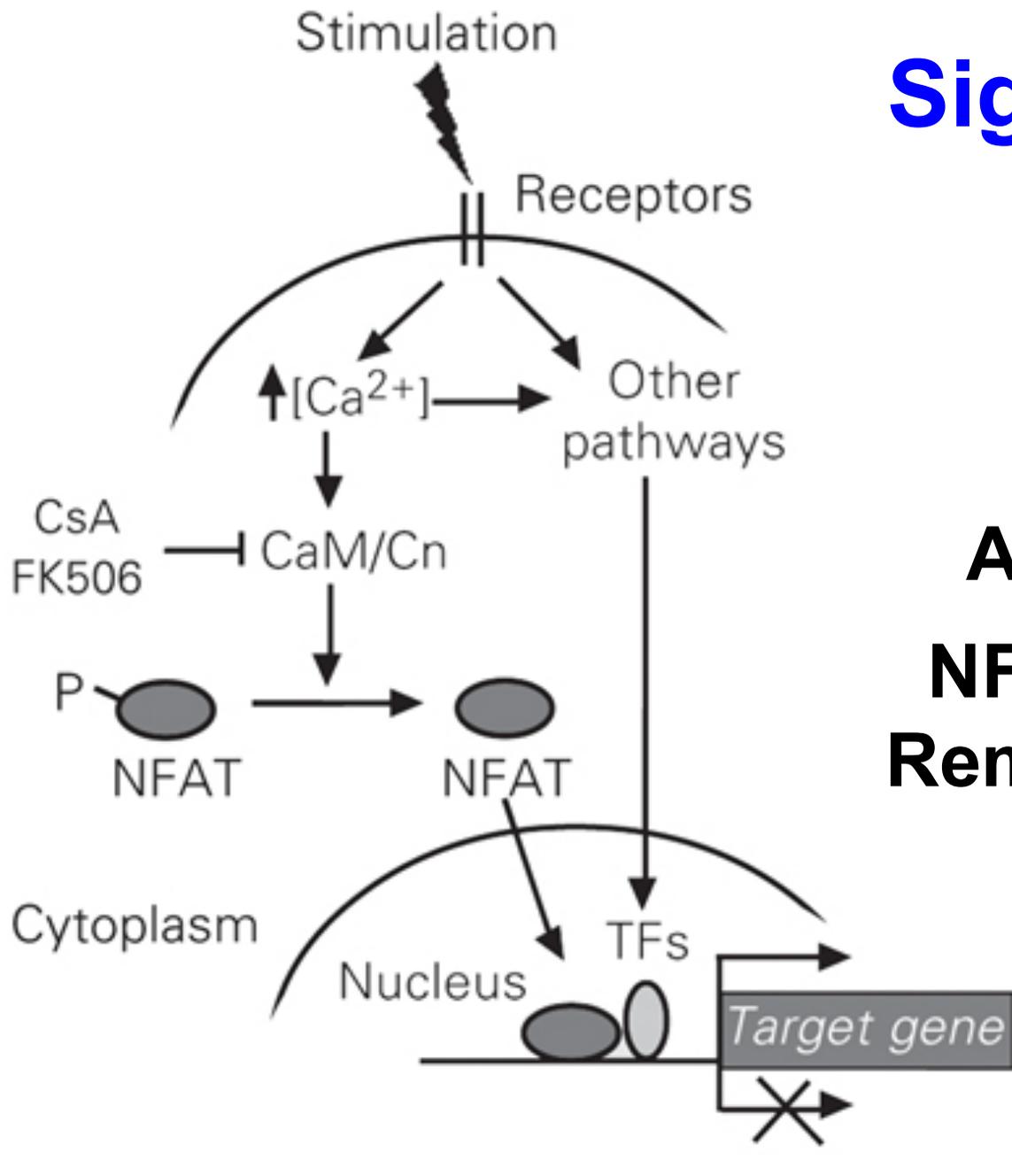
KEYWORDS	Proteins (number)	Families (number)	Length (Ave)	Z – Score
<u>Differentiation</u>	1406	422	439	18.8
Transcription	11223	1653	442	14.6
<u>Transcription Regulation</u>	9758	1554	413	14.3
<u>Spermatogenesis</u>	332	189	280	13.9
DNA Condensation	317	130	300	13.3
Cell Cycle	4278	612	494	12.2
mRNA Processing	1575	249	516	10.9
mRNA Splicing	716	180	459	10.1
Mitosis	718	215	620	9.4
<u>Apoptosis</u>	810	211	465	9.4

Xie H, et al., *J. Proteome Res* 6: 1882-1932 (2007)

Functions of Structured Proteins vs. IDPs

- **Sequence → Structure → Function ($Z < -1$)**
 - Catalysis,
 - Membrane transport,
 - Binding with DNA, RNA, Proteins, IDPs & molecules
- **Sequence → IDP Ensemble → Function ($Z > +1$)**
 - Signaling, Dunker AK, et al., *Biochemistry* 41: 6573-6582 (2002)
 - Regulation, Dunker AK, et al., *Adv. Prot. Chem.* 62: 25-49 (2002)
 - Recognition, Xie H, et al., *Proteome Res.* 6: 1882-1898 (2007)
 - Control. Vucetic, S. et al., *Proteome Res* 6: 1899-1916 (2007)
Xie H, et al., *Proteome Res* 6: 1917-1932 (2007)

Signaling Pathway



Calmodulin (CaM)
Calcineurin (Cn)
Nuclear Factor of Activated T- Cells (NFAT)

NFAT-poly-P in an IDP tail.
Remove Ps, activates NLS

- NFAT → nucleus**
- turns on genes**
- T-cells activated**
- reject transplant**

Nuclear Factor of Activated T-cells (NFAT)

Transcription Factor (TF) Family

NFAT: Phosphorylation → Inactivation

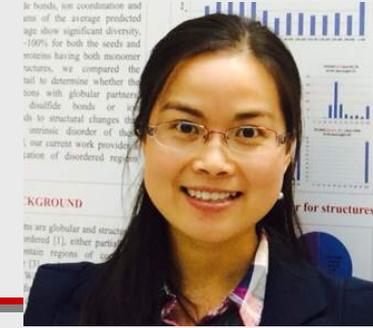
Ca²⁺/CaM → CaN Activation

Plays key roles in the following biological processes:

- **T-cell Activation**
- **Myocardial development**
- **Cancer metastasis**
- **And many more**
- **Angiogenesis**
- **Skeletal muscle development**

Pan MG et al., *Curr Mol Med* 13:543-554 (2013).

NFAT Family of TFs`

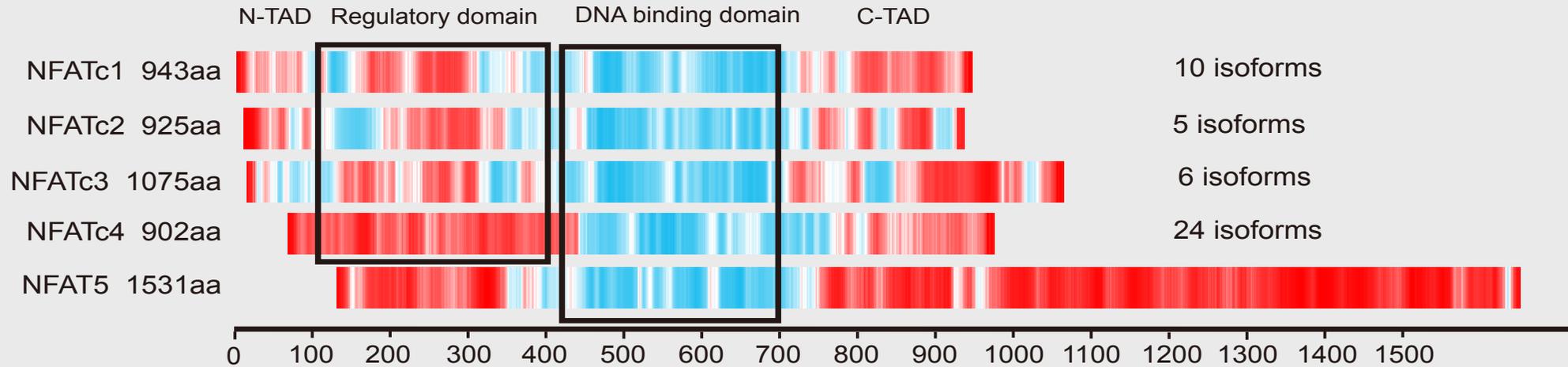


Jianhong Zhou

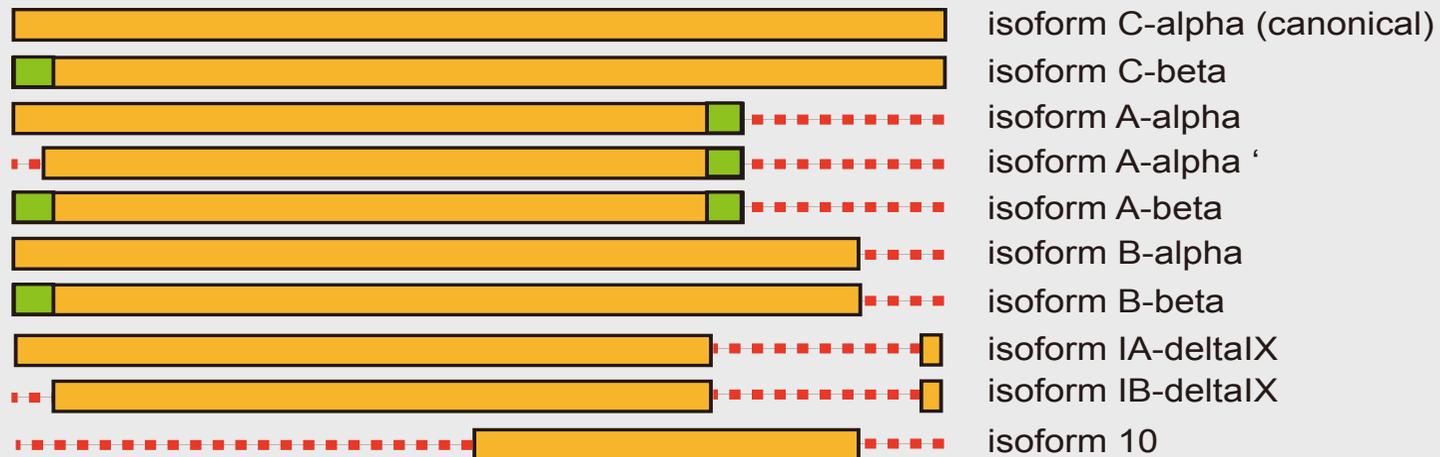


Suwen Zhao

A. Disorder Prediction

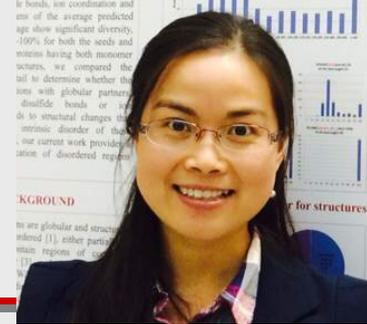


B. Splice Variants of NFATc1



**Zhou J et al.,
J Mol Biol
 430: 2342-
 2359 (2008)**

NFAT Family of TFs`



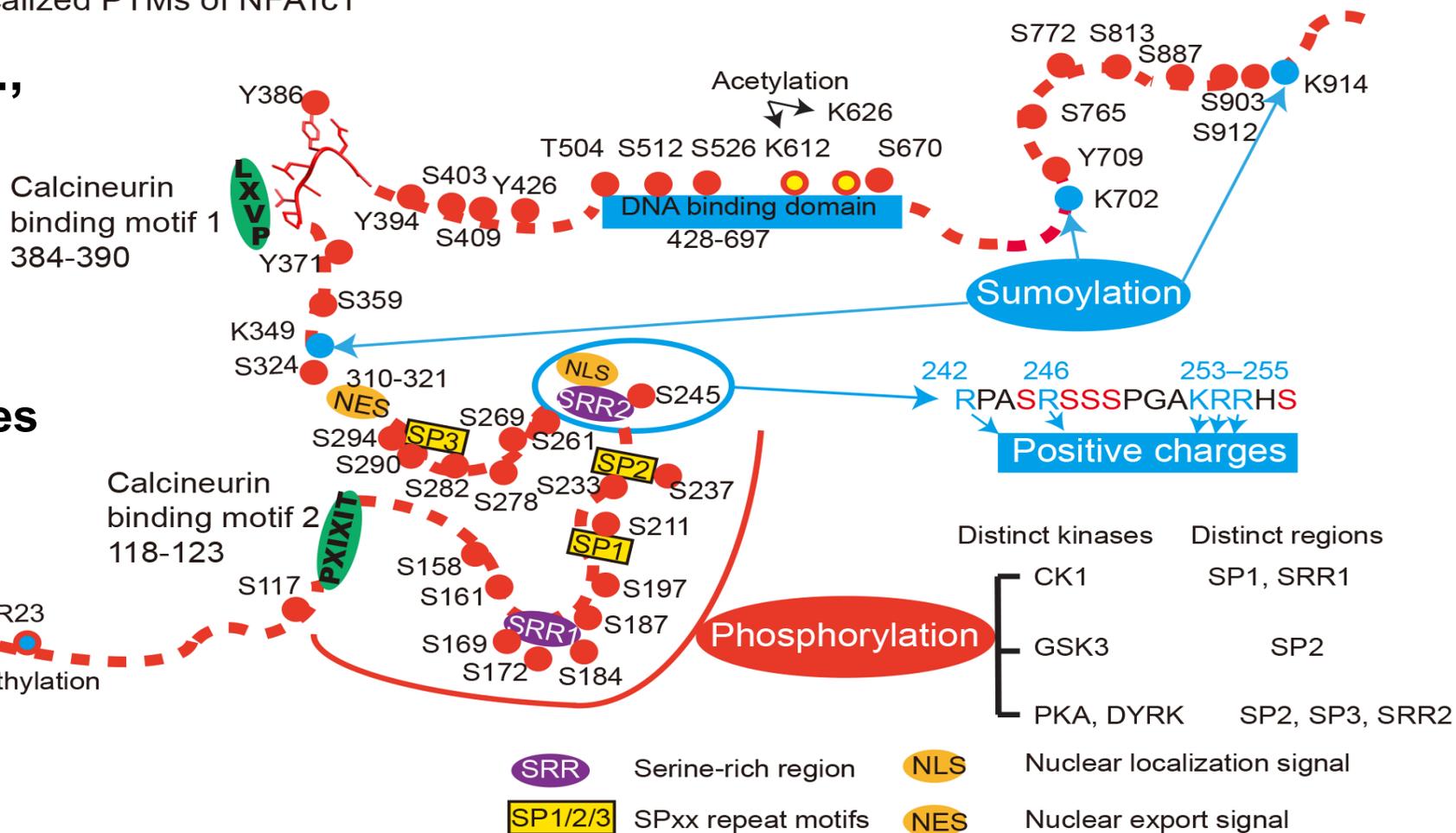
Jianhong Zhou



Suwen Zhao

C. Multiple IDR-localized PTMs of NFATc1

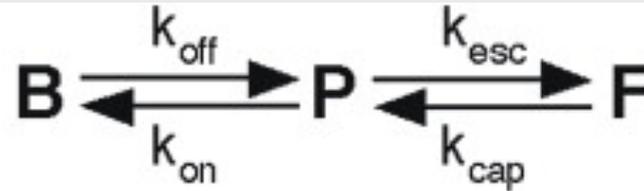
Zhou J et al.,
J Mol Biol
430: 2342-2359 (2018)



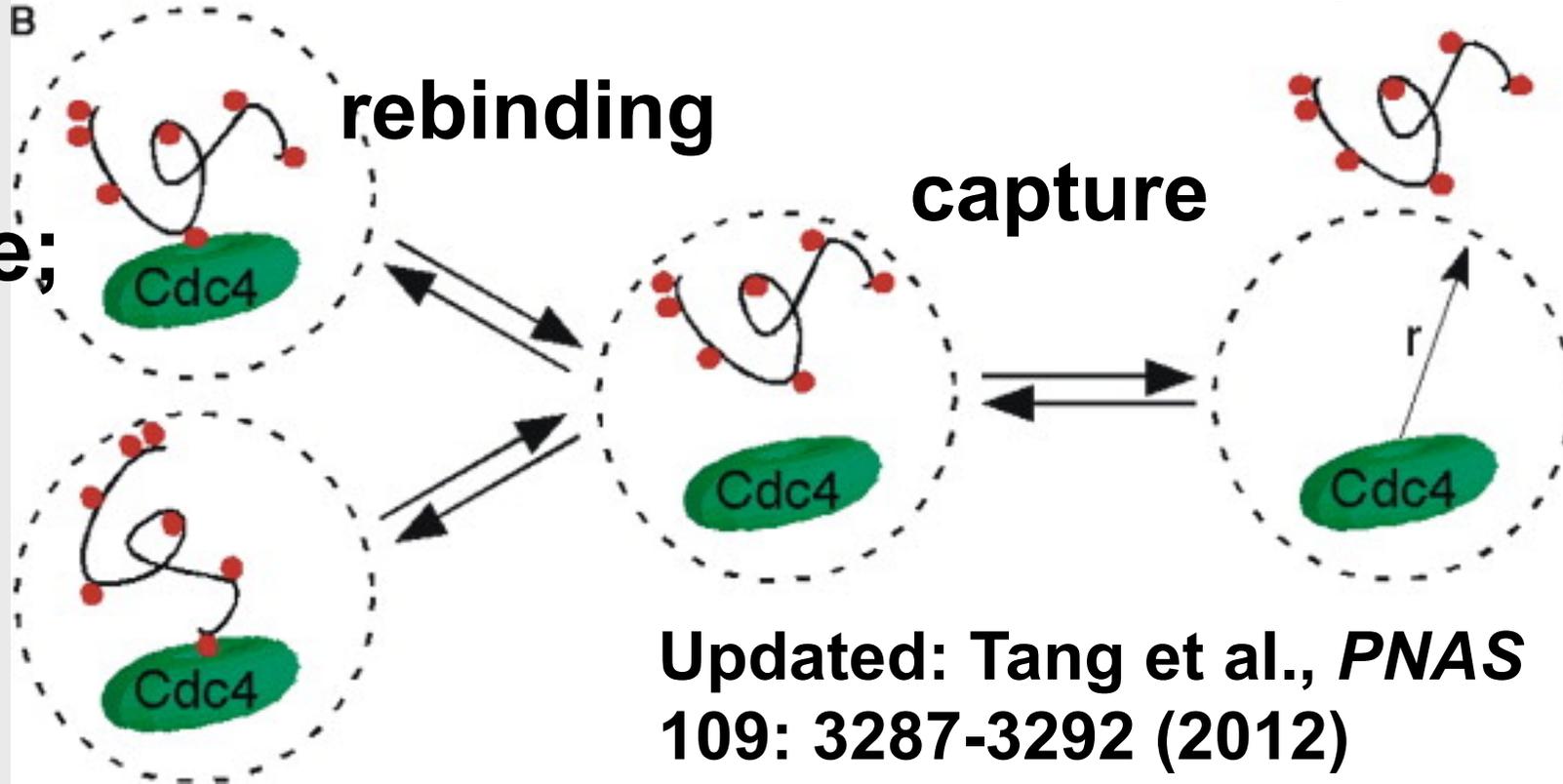
Cdc4-Sic1 & NFAT-NLS: ON-OFF Switches From Multiple Phosphates in IDP Regions

Overall Idea:

If # phosphates under threshold, then escape;
If over threshold, then rebinding.



Klein et al., *Curr Biol* 13: 1669-1678 (2003)



Many Proteins have **PTM Clusters**

Proteins	Suggested Concept	Reference
Histones	Histone Code	Strahl BD & Allis CD Nature 403:41-45 (2000)
p53, tubulin, Cdc25c, RNAP II	Molecular Barcode	Yang JX Oncogene 24: 1653-1662 (2005)
Transcription Factors	PTM Code	Benavoun BA & Veitia, Trends Cell Biol 19:189-197 (2009)
Various	Combinatorial PTMs	Lothrop AP et al., FEBS Lett 587:1247-1257 (2013)
P300 / CBP	Coactivator Code	Gamble MJ & Freedman LP, TIBS 27:165-167 (2002)
RNAP II CTD	Hyper-/Hypo-phosphorylation	Xu YX et al., Genes & Dev 17: 2765-2776 (2003)
Forkhead Box	FoxO Code	Calnan DR & Brunet A, Oncogene 27: 2276-2288 (2008)
p53	Cooperative Integrators	Meek DW & Anderson CW CSH Perspect Biol 1: a000950 (2009)

Cited in Pejaver et al., Protein Sci 23: 1077-1093 (2014)

PTM Clusters → PTM Codes

- For **PTM clusters** in Histones, p53, tubulin, Cdc25c, FoxO, RNAP II CTD, etc., the concept is that **different PTM patterns** lead to **different signaling consequences**.
- Thus, a **Histone** or **PTM code** likely exists.
- Predictions & experiments show that these **PTM clusters** are located in **IDP regions**.
- Bioinformatics extensions suggest that **PTM clusters** in **IDP regions** are very common.
- Thus, **PTM codes** are almost certainly very widely used for modulating **cell signaling**.



Pedja Radivojac



Vikas Pejaver

PTM Codes: Located in IDP Regions **Modulated by AS, Thus IDP-AS-PTM**

Proteins	Code Name	HITS	+AS
Histones	<i>Histone Code</i>	44,260	184
CREB BP	<i>Coactivator Code</i>	10,305	89
RNA Polymerase II	<i>Hyper/Hypo Phos.</i>	30,008	615
p53	<i>Molec. Barcode</i>	87,167	532
Tubulin		29,998	92
Forkhead Box	<i>FOXO Code</i>	2,357	6
Forkhead Box 1	<i>PTM Code</i>	3,372	8
Forkhead Box 4		336	4

References for **PTM Codes**:

New Idea: PTM Codes Modulated by AS

Histone Code – Strahl BD & Allis CD. *Nature*
403:41-45 (2000)

Coactivator Code – Gamble MJ & Freedman LP: *TIBS*
27:165-167 (2002)

Hyper/Hypo Phos – Xu YX et al., *Genes Dev*
17:2765-2776 (2003)

Molecular Barcode – Yang XJ *Oncogene* 24:1653-1662
(2005)

FOXO Code – Calnan DR & Brunet A: *Oncogene*
27:2276-2288 (2008)

PTM Code – Benayoun BA & Veitia RA: *Trends Cell Biol*
19:189-197 (2009).

The **IDP**–**AS**–**PTM** Toolkit Hypothesis

IDP, **AS**, & **PTM** shown to collaborate to yield complex signaling for the following proteins:

- NFAT family – transcription factors
- GPCR family – membrane signaling proteins
- Sarc Kinase family – signaling enzymes

Many proteins associated with cancer, cellular differentiation, conversion to stem cells, and so on all contain **IDPs**, **AS**, and **PTMs**, suggesting that this toolkit perhaps used by all these proteins. Have not yet shown their co-localization and collaboration – for the future.

Zhou J et al., J Mol Biol 430: 2342-2359 (2018)

Key Functions for the Evolution of Complex Multicellular Organisms

Complex multicellular organisms require the following:

- Cell adhesion;
 - Communication between cells;
 - Developmental programs;
 - Regulation of the developmental programs;
 - Cell-specific biochemistry.
- Nicklas & Newman
Evol Devel Biol
15: 41-52 (2013)

IDPs, **AS**, & **PTMs** common (universal?) among proteins that are involved in **all of these functions!!**

Dunker *AK et al.*, *Semin Cell Devel Biol* 37: 44-55 (2015)

IDPs and Gene Regulation



Bin Xue

Shinya Yamanaka (2012 Nobel Prize)

Overexpress 4 transcription factors (**TFs**):

All 4 of these **TFs** very rich in predicted **IDP AAs**:

Sox2 (**100%**), Oct4 (**67%**), Klf4 (**97%**), c-Myc (**80%**)

Adult fibroblast cells → induced Pluripotent Stem Cells (iPSCs)

The key **TFs** identified by >10 years of **trial and error** from a large number of additional **TFs**. Many **TFs** help with transdifferentiation by improving efficiency. Most of these **TFs** are rich in **predicted disorder**.

Xue B *et al.*, *Mol BioSys* 8:134-150 (2012)

IDPs and Gene Regulation



Julian Gough

Morgrify: An Algorithm (<http://morgrify.net>)

Input: **gene expression data** for different **cell types & known regulatory networks;**
data for 173 cell types, 134 tissues

Output: **Atlas of transcription factor sets:**
(**any cell type A**) → (**any cell type B**)

Results: Predicts **TF sets** for 5 known transdifferentiations
Predicts **TF sets** for 2 new transdifferentiations
Experiments worked on first try in both cases!!

Rackham OJL *et al.*, *Nature Genetics* 48: 331-335 (2016)

(seminar link: https://www.dropbox.com/s/5rf7s4cfkzrlwu9/CSHL-Asia_2018.pptx?dl=0)

IDPs and Gene Regulation

Rackham OJL *et al.*, *Nature Genetics* 48: 331-335 (2016)

Kamaraj US *et al.*, *Cell Cycle* 15: 3343-3354 (2016)



Julian Gough

ESC – Embryonic Stem Cell

MSC – Mesenchymal Stem

Cells

Since 2016

Previously known Transformations

1. Fibroblasts → Myoblasts (1998)
2. B-cells → Macrophages (2004)
3. Fibroblasts → iPSCs (2007)
4. Fibroblasts → Hepatocytes (2011)
5. Fibroblasts → Heart (2013)

Predicted & Confirmed

1. Fibroblasts → Keratinocytes
2. Keratinocytes → epithelial cells

1. ESC → Endothelial Cell
2. iPSC → Endothelial Cell
3. Fibroblast → Endothelial
4. Fibroblast → Astrocyte
5. ESC → Astrocyte
6. iPSC → Astrocyte
7. MSC → Astrocyte
8. ESC → Keratinocyte
9. iPSC → Keratinocyte
- + 2 more, All of first try!

Summary

- **Sequence → Structure → Function**

- Catalysis,
- Membrane transport,
- Binding with DNA, RNA, Proteins, **IDPs** & molecules

- **Sequence → IDP Ensemble → Function**

- **Signaling**, Dunker AK, et al., *Biochemistry* 41: 6573-6582 (2002)
- **Regulation**, Dunker AK, et al., *Adv. Prot. Chem.* 62: 25-49 (2002)
- **Recognition**, Xie H, et al., *Proteome Res.* 6: 1882-1898 (2007)
- **Control**.
Vucetic, S. et al., *Proteome Res* 6: 1899-1916 (2007)
Xie H, et al., *Proteome Res* 6: 1917-1932 (2007)

A STRUCTURE-BASED Toolkit

Active Site



A rock-like **structured protein**



Substrate

Product



Lock and key,
induced fit;
many proteins,
many functions

The **IDP**-**AS**-**PTM** Developmental Toolkit



An **IDP** or **IDP Region**,
+ **PTMs**
+ **AS**

One IDP, many shapes,
many functions,
provides a toolkit for
complex signaling &
cellular differentiation

Intrinsically Disordered Proteins

THANK YOU!!!

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